The Th2/Th17 pathway in asthma and the relevant clinical significance

Kalliopi Domvri¹, Georgios Tzimagiorgis², Despoina Papakosta¹

¹Asthma Clinic, Pulmonary Department, Aristotle University of Thessaloniki, George Papanikolaou Hospital, Exochi ²Laboratory of Biological Chemistry, School of Medicine, Faculty of Health Sciences, Aristotle University of Thessaloniki, Thessaloniki, Greece

Key words:

- Th2/Th17
- Asthma
- Phenotypes
- Biomarkers
- Treatment

Correspondence:

Despoina Papakosta, Professor of Pneumonology-Lung Immunology, Pulmonary Department Aristotle University of Thessaloniki, Greece e-mail: depapako@gmail.com

SUMMARY

Asthma is a heterogeneous chronic disease of the airways, characterized by different phenotypes. The principal pathophysiological pathway appears to be Th2 dependent eosinophilic inflammation mainly produced by T helper 2 (Th2) cells. More recently epithelial innate lymphoid cells (ILC2) cell shave been implicated as another source of Th2 cytokines leading to bronchial eosinophilia without previous allergen sensitization. Another pathogenic pathway is the non-Th2 type, mediated by Th1 and especially Th17 lymphocytes, responsible for neutrophilicin flammation. Furthermore, recent studies have associated Th-17 cells with allergic inflammation and eosinophilic asthma. Ongoing clinical trials are expected to further elucidate the role of different cells in the evolution of asthmatic inflammation and also the role of established or novel potential biomarkers in routine clinical practice aiming to maximize drug efficacy in asthmatics. In the present review, we summarize the above mentioned mechanisms focusing on T-helper cell subset plasticity which led to the identification of dual positive Th2/Th17 inflammation. Pneumon 2018, 31(3):174-182.

INTRODUCTION

Asthma is a heterogeneous disease of the airways characterized by airway inflammation and bronchial hyperresponsiveness (BHR) leading to reversible airway obstruction¹. This chronic disease affects many people, men and women, young and old, worldwide. It is defined by a history of respiratory symptoms (such as wheeze, shortness of breath, chest tightness and cough), which vary over time and in intensity². The majority of asthma patients are well controlled by conventional therapies such as inhaled corticosteroids. However, about 5-10% of asthma patients have a severe and complex condition, described as "fatal or near fatal asthma", "severe asthma", "steroid-dependent asthma", "steroid-insensitive asthma", or "irreversible asthma"³.

For over 20 years, asthma has been considered a Th2type dependent allergic disease, characterized by Th2 cells producing high levels of type 2 interleukins (ILs), such as IL-4, IL-5 and IL-13. Besides, other studies suggested that Th-1 cells producing interferon (IFN)- γ display a regulatory function in allergic asthma⁴. Although the Th-1/Th2 mechanism provided the initial framework for asthma management, the discovery of a distinct subpopulation of CD4+ T cells that produce IL-17A, IL-17F, IL-22, TNF- α , and IL-21 led to a major revision of the Th-1/Th2 hypothesis (Figure 1). Th17 cells are differentiated and activated by several cytokines such as transforming growth factor TGF β , IL-6 together with IL-21 and IL-23⁵.

In addition to Th2 and Th17 cells, the heterogeneity of asthmatic patients suggests that also other factors must be involved in regulating asthma inflammation. Indeed, recent studies have implicated innate lymphoid cells (ILCs) of non-T, non-B effector cells that are antigen-nonspecific, have conserved effector cell functions and play crucial roles in tissue homeostasis, repair and remodeling and in innate immunity to pathogenic and nonpathogenic microorganisms⁶. ILCs are classified into three categories (Type 1, Type 2 and Type 3 ILCs) depending on their ability to produce Th1, Th2 and Th17 cell-associated cytokines⁶. In specific, ILC type 2 (ILC2) have been associated with



Allergic inflammation

FIGURE 1. Type 2 Innate Lymphoid Cells and Th cell subsets involved in allergic and non-allergic inflammation in asthma.

asthma by producing a broad array of cytokines, including IL-5, IL-13 and IL-17⁷.

Asthma was initially categorized in terms of 'allergic' or "nonallergic" asthma. A distinction was then made when sputum became available between eosinophilic and non-eosinophilic asthma². The last decade, a global approach for the understanding of asthma pathogenesis has introduced the concept of phenotypes as a grouping of clinical/physiologic characteristics, triggering factors and inflammatory components². A new approach includes the addition of genetic or blood biomarker for the classification of disease entities within the asthma syndrome which led to the introduction of the term endotype³. Indeed, asthma endotyping has shed light into key pathogenic mechanisms for this complex disorder⁸.

Recent discoveries revealed possible subgroups of Th2 high asthma that differ in terms of both the presence of underlying allergy and the potential source of type 2 cytokines. The current concept involves Th2-high asthma, eosinophilic, characterized by high levels of type 2 interleukins (ILs), and involves type 2 helper T cells (Th2 cells), mast cells, basophils, B cells and ILC2s^{4,8,9}. The fact that ILC2 produce Th2 cytokines could explain severe eosinophilic inflammation, when classical Th2 mediated allergy is absent, which is further supported in other studies¹⁰.

On the other hand lays Th2-low/Non-Type 2 as non eosinophilic asthma, where Th17 cells are involved (IL-17A, IL-23, IL-22, IL-6), mostly characterized by neutrophilic inflammation¹¹. An interesting issue for Th2 and Th17 cells is the qualitative difference concerning their response to glucocorticoid treatment, as IL-17 production was shown to be less susceptible to inhibition by glucocorticoids when compared to IL-4 and IL-5 production¹².

According to recent literature possible endotypes associated with eosinophilic phenotypes include, early onset allergic asthma with or without obesity, aspirin sensitive asthma and late onset eosinophilic asthma, exacerbation prone asthma, and exercise induced asthma,whereas those associated with non-eosinophilic asthma and for which the pathobiologic pathways are not yet defined, include those patients with obesity-related late onset asthma, asthma with fixed airflow obstruction and very little inflammation (paucigranulocytic), and asthma associated with neutrophilia¹¹.

BIOMARKERS IN ASTHMA

A biomarker is defined as "a characteristic that is objectively measured and evaluated as an indicator of normal biological processes, pathogenic processes or pharmacological responses to a therapeutic intervention"9. Cytokines play a significant role in the pathogenesis and chronic inflammation of the airways in asthma and therefore, interleukins as biomarkers could identify endotypes and subtypes of asthma, reflecting the predominant pathophysiological mechanism¹³. The use of biomarkers can potentially help avoid unnecessary morbidity from high-dose corticosteroid therapy, and allow the most appropriate and cost-effective use of targeted therapies¹⁴. As cytokines include markers of inflammation, several studies have been trying to identify molecular endotypes based on serum cytokine profile¹⁵. However, the persistent airway inflammation in asthma is caused by a large variety of cytokines that recruit, activate and perpetuate the inflammatory cells in the asthmatic airways. As a result, several attempts to inhibit inflammatory cytokines in asthma with blocking antibodies have shown poor results so far and besides studies of targeted therapy in non-phenotyped asthma did not show profound or even any efficacy¹⁶. Furthermore, it is a fact that there is no consensus how to best identify asthma endotypes and what therapy to use for a given endotype. Apart from cytokine profile, the use of blood eosinophils can facilitate individualized treatment and management of asthma.

Th2-specific biomarkers identified so far include sputum/blood eosinophils, total serum IgE, the fraction of exhaled nitric oxide (FeNO) and bronchial epitheliumderived proteins such as periostin and DPP4¹¹. The first biomarker proposed to predict corticosteroid response was the eosinophil count in sputum and blood¹⁷. Furthermore, total serum IgE is used specifically to identify allergic asthma phenotype². However, IgE has a low sensitivity and correlates poorly with eosinophilic inflammation¹⁸. FeNO is also a biomarker of Th2 inflammation¹⁹ and is suggested to be used as a predictor of steroid responsiveness more consistently than other parameters²⁰. According to recent literature, in patients with mild to severe asthma, blood eosinophils have the highest accuracy in the identification of sputum eosinophilia when compared to serum periostin and exhaled nitric oxide (FeNO)²¹. Indeed, serum periostin, exhaled nitric oxide and blood eosinophil counts are the most promising biomarkers until now that could identify patients patients most likely to derive benefit from biologic agents targeting IgE, IL-5 and IL-4/13²².

The combined evaluation of FeNO and peripheral blood eosinophil counts represent two significant biomarkers of asthma, comparable to sputum eosinophil count and are often used to distinguish asthma phenotypes and even identify responders to inhaled corticosteroid treatment^{2,23}. An count of >400 eosinophils/ μ L is associated with more severe asthma²⁴.Concerning the non Th2 pathway biomarkers, data are limited compared to Th2 type. Sputum neutrophils, mixed granulocytic and paucigranulocytic patterns include the most commonly used biomarkers²⁵.

Th2 cytokines

Th2 cytokines (IL-4, IL-5, IL-9, IL-10, and IL-13) have a substantial effect on the pathogenesis of atopic diseases²⁶. It is currently suggested that apart from Th2 cells, ILC2s are also responsible for the production of the majority of Th2 cytokines in the airway²⁷.

IL-4/IL-13

Th2 cytokines IL-4 and IL-13 share significant pathways and many biological activities concerning asthma. In specific, they play an important role in the identification of the presence of eosinophilic inflammation and also are key factors in IgE synthesis by B cells, mucus production, bronchial fibrosis and airway hyperresponsiveness in asthma^{28,29}. There is a plethora of studies associating IL-4, IL-13 and asthma. When comparison was made between asthmatics and healthy controls, levels of IL-mRNA, protein levels in serum, bronchoalveolar lavage fluid (BAL), bronchial biopsies and exhaled breath condensate were found higher in asthmatics³⁰⁻³⁴. Similarly in other studies, increased IL-13 mRNA and protein levels are also found in sputum, BAL and bronchial biopsies of patients with asthma when compared to controls³⁵⁻³⁷.

Furthermore, it is well-known that single-nucleotide polymorphisms (SNP) can be used to assess genetic disorders. A recent meta-analysis showed that IL-4 C-589T and C-33T were associated with asthma in Europeans³⁸. As for IL-13, two SNPs, positioned at regions +2044G/A and +1923C/T have been suggested to play critical role in the development of asthma³⁹⁻⁴¹.

Anti-IL-13 and anti-IL-13/anti-IL-4 receptors targeted therapies with humanized monoclonal antibodies are currenlty used as add-on therapy in patients with Th2-high inflammation with uncontrolled asthma despite maximum therapy. Specific studies have shown that Th2-high and especially periostin-high groups of asthmatics with moderate-to-severe uncontrolled asthma are suggested to compose the group that could benefit from anti-IL-13 therapy⁴². Anti-IL-13 biologic agents (anrukinzumab, lebrikizunab and tralokinumab), are currently under clinical evaluation to further elucidate the use of predictive Th-2 biomarkers⁴³.

Besides, IL-4 and IL-13 share some structural similarities and they bind the IL-4Ra/IL-13Ra1 receptor complex by which the transcription factor STAT-6 is activated⁴⁴. Based on this concept, biologic agent such as dupilumab that target the dual cytokines IL-4/13 may be more encouraging approach for those patients suffering from refractive difficult-to-control eosinophilic asthma. Indeed, recently Wenzel et al reported that dupilumab increased lung function and reduced severe exacerbations in patients with uncontrolled persistent asthma irrespective of baseline eosinophil count and had a favourable safety profile⁴⁵. More recently, another study reported that in patients with glucocorticoid-dependent severe asthma, dupilumab treatment reduced the rate of severe exacerbations and the use of oral steroids along with increasing the FEV_1^{46} . Significantly lower rates of severe asthma exacerbations, as well as better lung function and asthma control were also reported in another study in patients treated with dupilumab47.

IL-5

IL-5 is produced by CD4 + Th2 lymphocytes and ILC2 cells and, to a lesser extent, also by natural killer T (NKT) cells, mast cells, and eosinophils themselves⁴⁸. Multiple studies have demonstrated the link between IL-5 and asthma as higher IL-5 mRNA levels⁴⁹ and IL-5 levels in sputum⁵⁰ have been associated with increasing eosinophil production or acute asthmatic exacerbations⁵¹. As for genetics, IL-5 C-746T was found to influence atopic outcomes⁵². Based on the above, IL-5 has been considered a suitable target for add-on biological therapies of severe eosinophilic asthma^{53,54}. In particular, the anti-IL-5 antibodies developed, include mepolizumab and reslizumab, and the IL-5 receptor antagonist benralizumab which are evaluated for the treatment of refractory eosinophilic asthma⁵⁵⁻⁵⁷. Apart from mepolizumab and reslizumab, already FDA approved⁵⁵, benralizumab has also recently obtained the approval of FDA on the basis of several successful randomized controlled trials⁵⁸.

IL-17A/IL-23

A distinct subpopulation of CD4+T cells produce Th-17 cells that by secreting IL-17, orchestrate the recruitment of neutrophil granulocytes in the lungs⁵⁹. Furthermore, IL-17 contribute to the development of airway fibrosis during asthma by enhancing the production of profibrotic

cytokines, proangiogenic factors, and collagen⁶⁰. Besides, in vitro studies investigating the role of epithelial-mesenchymal transition (EMT) in asthma have shown IL-17 synergization with IL-4 and TGF- β promotes EMT with the expression of mesenchymal markers^{61,62}.

Th17 cells produce IL-17A, IL-17F, IL-21 and IL-22 cytokines⁵⁹. IL-17A has been involved in severe asthma characterized by airway intense neutrophil infiltration and less responsive to corticosteroids^{59,63}. Furthermore, these steroid-insensitive patients could be classified as a Th2 low phenotype of asthma². Clinical studies have shown that levels of IL-17A is sputum, bronchoalveolar lavage fluid (BALF) and serum of asthmatic patients are significant higher in asthmatics than in healthy subjects and that these are correlated with the severity of disease⁶⁴. Additionally, in a study on pre-school children with history and physical exam in favor of asthma which cannot be tested by spirometry, they concluded that IL-23 serum levels might be an auxiliary biomarker for the diagnosis of asthma⁶⁵.

Recently, Fattahi et al showed that atopy is associated with lower numbers of IL-17 cells in asthmatic airways⁶⁶. Current data suggest that IL-17 has also been implicated in Th2 cell-mediated eosinophilic airway inflammation in mouse models of asthma⁶⁷ along with increased levels of IL-2368 or in asthmatic patients with allergy after a challenge with house dust mite⁶⁹. Furthermore, recently Camargo et al reported that inhibition of IL-17 even in exacerbated asthmatic patients significantly contributed to the control of Th1/Th2/Th17 inflammation, chemokine expression, extracellular matrix remodeling, and oxidative stress in a murine experimental asthma model exacerbated by Lipopolysaccharide (LPS)⁷⁰. As for genetics, a recent meta-analysis concerning the association between IL-17A polymorphisms and asthma risk suggested that the IL-17A -737C/T polymorphism provides protection against the disease, whereas the IL-17A -197G/A polymorphism does not contribute to asthma risk⁷¹.

However, so far, results from clinical trials targeting IL-17 Receptor (Brodalumab) including moderate to severe asthmatics, showed no improvement in asthma outcomes. This fact could be apparently attributed to inadequate selection of patients with asthma⁷². More specifically, although there was no effect of brodalumab on the primary outcome (the Asthma Control Questionnaire score), researchers, based on a subgroup analysis, suggested a new phenotype including patients with high reversibility of FEV1 in response to albuterol and a new

endotype which is IL-17R-dependent⁶⁰. In future trials targeting IL-17 pathway, the selection of patients based on sputum neutrophilia could exclude Th2 high asthmatics that are less likely to respond to an IL-17-targeted therapy.

Dual positive Th2/Th17 cells

Athough T-helper cells were thought to be fully differentiated, expressing a master regulatory transcription factor and their development from naive CD4 cells was considered to be lineage specific⁷³, Cosmi et al demonstrated that there is great plasticity in human Th17 cells even toward the Th2 phenotype, suggesting the existence of CD4+T cells able to produce both Th17 related (IL-17A) and Th2 (IL-4) related cytokines⁷⁴. Besides, several in vitro studies and animal studies suggest that T-helper cell subsets display plasticity by changing their transcription factor or by expressing multiple transcription factors⁷⁵⁻⁷⁹.

Asthmatic patients can suffer from a predominant eosinophilic inflammation usually seen in mild-to-moderate disease, from neutrophilic inflammation in more severe disease or even mixed eosinophilic/neutrophilic inflammatory response⁸⁰. The underlying T cell response is predominated by Th2, Th17, or a mixed Th2/Th17 cell immune response.

A recent study showed that asthma is associated with a higher frequency of dual-positive Th2/Th17 cells in BAL fluid¹². They concluded that Th2/Th17 (predominant) subgroup of asthmatic patients manifested glucocorticoid resistance in vitro and also had the greatest airway obstruction and hyperreactivity compared with the Th2 (predominant) and Th2/Th17 (low) subgroups. Moreover, in experimental animal models, IL-17 has not only been involved to produce airway intense neutrophil infiltration but to exacerbate Th2 cell mediated eosinophilic airway inflammation and hyperresponsiveness^{67,68}.

However, in another study, Choy et al investigated the potential of Th2 cytokine suppression in promoting TH17 responses in a preclinical model of allergen-induced asthma and concluded that IL-13 and IL-17A reciprocally regulate the expression of their target pathways in the lung⁸¹. In specific, IL-13 stimulation repressed the expression of the Th17 genes, with a trend for a similar repressive effect of IL-17A stimulation on Th2 genes.

Furthermore, in an effort to discover the possible mechanism of severe late-onset hypereosinophilic phenotype, ILC2 activation along with dual positive Th2/Th17 inflammation has been proposed¹¹. Besides, recent data suggest that combination therapies targeting both pathways may maximize therapeutic efficacy across a patient population comprising both Th2 and Th17 endotypes¹².

CONCLUSIONS AND FUTURE PERSPECTIVES

As new research data concerning the different T helper lymphocyte pathogenic pathways are being developed in the asthmatic inflammation cascade, new effective individualized therapies in severe asthma are urgently needed to block specific inflammatory pathways using monoclonal antibodies. The only approved therapies so far include anti-IL-5 IgG (Mepolizumab, Reslizumab, Benralizumab) for severe eosinophilic asthma and anti-IgE (Omalizumab) for the treatment of severe allergic asthma. There is none approved biomarker for Non-Type 2/Th2 low asthma. One could possibly further suggest that combination therapies targeting both pathways might maximize therapeutic efficacy across a patient population comprising both Th2 and Th17 endotypes. Clinically, the ability to identify a Th2 or Th17 high or a mixed asthma phenotype on the basis of testing blood eosinophils may facilitate the use of effective biologically targeted approaches in asthmatic patients. The fact that analyzing the cytokine pattern in serum samples does not give us information on the source of the assessed cytokine may reflect the significance of local environment like lung tissues or intracellular molecular methods in analyzing the cytokine levels.But meanwhile, the feasibility of serum sampling as a noninvasive method to analyze cytokine levels has directed the interests toward noninvasive methods rather than invasive ones.

ΠΕΡΙΛΗΨΗ

Το μονοπάτι Th2/Th17 στο άσθμα και η κλινική του σημασία

Καλλιόπη Δόμβρη¹, Γεώργιος Τζημαγιώργης², Δέσποινα Παπακώστα¹

¹Πνευμονολογική Κλινική, Αριστοτέλειο Πανεπιστήμιο Θεσσαλονίκης, Γ.Ν.Θ. «Γ. Παπανικολάου», Θεσσαλονίκη, ²Εργαστήριο Βιολογικής Χημείας, Σχολή Επιστημών Υγείας, Τμήμα Ιατρικής, Αριστοτέλειο Πανεπιστήμιο Θεσσαλονίκης

Το άσθμα είναι μια ετερογενής χρόνια νόσος των αεραγωγών, που χαρακτηρίζεται από διαφορετικούς φαινοτύπους. Η κύρια παθοφυσιολογική οδός είναι εξαρτώμενη από την Th2 ηωσινοφιλική φλεγμονή που παράγεται κυρίως από βοηθητικά T2 λεμφοκύτταρα (Th2). Πρόσφατες βιβλιογραφικές αναφορές προτείvouv τα μη ευαισθητοποιημένα λεμφοκύτταρα του βρογχικού επιθηλίου (innate lymphoid cells, ILC₂) ως μία άλλη πηγή Th2 κυτταροκινών, με αποτέλεσμα την παραγωγή ηωσινοφίλων χωρίς προηγούμενη αντιyovική ευαισθητοποίηση. Άλλο παθοφυσιολογικό μονοπάτι είναι η μη Th2 φλεγμονή που εξελίσσεται μέσω των Th1 λεμφοκυττάρων και των Th17 που εμπλέκονται στην ουδετεροφιλική φλεγμονή. Όμως, πρόσφατες μελέτες έχουν συσχετίσει και τα Th17 κύτταρα με την αλλεργική φλεγμονή και το ηωσινοφιλικό άσθμα. Οι τρέχουσες κλινικές μελέτες αναμένεται να διευκρινίσουν περαιτέρω τον ρόλο των διαφόρων κυττάρων στην εξέλιξη της ασθματικής φλεγμονής και επίσης τον ρόλο των καθιερωμένων ή νέων πιθανών βιοδεικτών στην καθημερινή κλινική πρακτική με στόχο τη μεγιστοποίηση της αποτελεσματικότητας των αντιασθματικών φαρμάκων. Στην παρούσα επισκόπηση, συνοψίζουμε τους προαναφερθέντες μηχανισμούς, επικεντρώνοντας στην πλαστικότητα κι ευελιξία των υποπληθυσμών των Τ-βοηθητικών κυττάρων και στην αναγνώριση της διπλής Th2/Th17 θετικής φλεγμονής.

Πνεύμων 2018, 31(3):174-182.

Λέξεις - Κλειδιά: Th2/Th17, Άσθμα, Φαινότυποι, Βιοδείκτες, Θεραπεία

REFERENCES

- 1. From the Global Strategy for Asthma Management and Prevention, Global Initiative for Asthma (GINA). 2015.
- 2. Wenzel SE. Asthma: defining of the persistent adult phenotypes. Lancet 2006;368(9537):804-13.
- Lotvall J, Akdis CA, Bacharier LB, et al. Asthma endotypes: a new approach to classification of disease entities within the asthma syndrome. The Journal of Allergy and Clinical Immunology 2011;127:355-60.
- 4. Afshar R, Medoff BD, Luster AD. Allergic asthma: a tale of many T cells. Clin Exp Allergy 2008;38:1847-57.
- Zhou L, Ivanov II, Spolski R, et al. IL-6 programs T(H)-17 cell differentiation by promoting sequential engagement of the IL-21 and IL-23 pathways. Nature Immunology 2007;8:967-74.
- Yu S, Kim HY, Chang YJ, DeKruyff RH, Umetsu DT. Innate lymphoid cells and asthma. The Journal of Allergy and Clinical Immunology 2014;133:943-50; quiz 51. Epub 2014/04/01.
- Spits H, Artis D, Colonna M, et al. Innate lymphoid cells--a proposal for uniform nomenclature. Nature Reviews Immunology 2013;13:145-9. Epub 2013/01/26.
- 8. Anderson GP. Endotyping asthma: new insights into key pathogenic mechanisms in a complex, heterogeneous disease. Lancet 2008;372(9643):1107-19.
- 9. Fajt ML, Wenzel SE. Biologic therapy in asthma: entering the

new age of personalized medicine. The Journal of asthma: Official Journal of the Association for the Care of Asthma 2014;51:669-76.

- Brusselle GG, Maes T, Bracke KR. Eosinophils in the spotlight: Eosinophilic airway inflammation in nonallergic asthma. Nature Medicine 2013;19:977-9. Epub 2013/08/08.
- Carr TF, Zeki AA, Kraft M. Eosinophilic and Noneosinophilic Asthma. American Journal of Respiratory and Critical Care Medicine 2018;197:22-37. Epub 2017/09/15.
- Irvin C, Zafar I, Good J, et al. Increased frequency of dual-positive TH2/TH17 cells in bronchoalveolar lavage fluid characterizes a population of patients with severe asthma. The Journal of Allergy and Clinical Immunology 2014;134:1175-86 e7.
- Kawaguchi M, Adachi M, Oda N, Kokubu F, Huang SK. IL-17 cytokine family. The Journal of Allergy and Clinical Immunology 2004;114:1265-73; quiz 74.
- Bartminski G, Crossley M, Turcanu V. Novel biomarkers for asthma stratification and personalized therapy. Expert Rev Mol Diagn 2015;15:415-30. Epub 2014/12/06.
- Liang Z, Liu L, Zhao H, et al. A Systemic inflammatory endotype of asthma with more severe disease identified by unbiased clustering of the serum cytokine profile. Medicine 2016;95(25):e3774.
- Heck S, Nguyen J, Le DD, Bals R, Dinh QT. Pharmacological Therapy of Bronchial Asthma: The Role of Biologicals. Interna-

tional Archives of Allergy and Immunology 2015;168:241-52.

- Woodruff PG, Modrek B, Choy DF, et al. T-helper type 2-driven inflammation defines major subphenotypes of asthma. American Journal of Respiratory and Critical Care Medicine 2009;180:388-95. Epub 2009/06/02.
- Jia G, Erickson RW, Choy DF, et al. Periostin is a systemic biomarker of eosinophilic airway inflammation in asthmatic patients. The Journal of Allergy and Clinical Immunology 2012;130:647-54 e10. Epub 2012/08/04.
- Chibana K, Trudeau JB, Mustovich AT, et al. IL-13 induced increases in nitrite levels are primarily driven by increases in inducible nitric oxide synthase as compared with effects on arginases in human primary bronchial epithelial cells. Clin Exp Allergy 2008;38:936-46. Epub 2008/04/04.
- Smith AD, Cowan JO, Brassett KP, et al. Exhaled nitric oxide: a predictor of steroid response. American Journal of Respiratory and Critical Care Medicine 2005;172:453-9. Epub 2005/05/20.
- 21. Wagener AH, de Nijs SB, Lutter R, et al. External validation of blood eosinophils, FE (NO) and serum periostin as surrogates for sputum eosinophils in asthma. Thorax 2015;70:115-20.
- 22. Arron JR, Choy DF, Scheerens H, Matthews JG. Noninvasive biomarkers that predict treatment benefit from biologic therapies in asthma. Annals of the American Thoracic Society 2013;10(Suppl):S206-13.
- Westerhof GA, Korevaar DA, Amelink M, et al. Biomarkers to identify sputum eosinophilia in different adult asthma phenotypes. The European Respiratory Journal 2015;46:688-96. Epub 2015/06/27.
- 24. Price DB, Rigazio A, Campbell JD, et al. Blood eosinophil count and prospective annual asthma disease burden: a UK cohort study. The Lancet Respiratory Medicine 2015;3:849-58. Epub 2015/10/24.
- Baos S, Calzada D, Cremades-Jimeno L, et al. Nonallergic Asthma and Its Severity: Biomarkers for Its Discrimination in Peripheral Samples. Frontiers in Immunology 2018;9:1416. Epub 2018/07/07.
- 26. Bellanti JA. Cytokines and allergic diseases: clinical aspects. Allergy and Asthma Proceedings 1998;19:337-41.
- Mirchandani AS, Besnard AG, Yip E, et al. Type 2 innate lymphoid cells drive CD4+ Th2 cell responses. Journal of Immunology 2014;192:2442-8.
- Wills-Karp M, Luyimbazi J, Xu X, et al. Interleukin-13: central mediator of allergic asthma. Science 1998;282(5397):2258-61.
- Beghe B, Barton S, Rorke S, et al. Polymorphisms in the interleukin-4 and interleukin-4 receptor alpha chain genes confer susceptibility to asthma and atopy in a Caucasian population. Clin Exp Allergy 2003;33:1111-7.
- 30. Walker C, Bauer W, Braun RK, et al. Activated T cells and cytokines in bronchoalveolar lavages from patients with various lung diseases associated with eosinophilia. American Journal of Respiratory and Critical Care Medicine 1994;150:1038-48. Epub 1994/10/01.
- 31. Humbert M, Durham SR, Ying S, et al. IL-4 and IL-5 mRNA and protein in bronchial biopsies from patients with atopic and nonatopic asthma: evidence against "intrinsic" asthma being

a distinct immunopathologic entity. American Journal of Respiratory and Critical Care Medicine 1996;154:1497-504. Epub 1996/11/01.

- 32. Ying S, Durham SR, Corrigan CJ, Hamid Q, Kay AB. Phenotype of cells expressing mRNA for TH2-type (interleukin 4 and interleukin 5) and TH1-type (interleukin 2 and interferon gamma) cytokines in bronchoalveolar lavage and bronchial biopsies from atopic asthmatic and normal control subjects. Am J Respir Cell Mol Biol 1995;12:477-87. Epub 1995/05/01.
- 33. Ying S, Humbert M, Barkans J, et al. Expression of IL-4 and IL-5 mRNA and protein product by CD4+ and CD8+ T cells, eosinophils, and mast cells in bronchial biopsies obtained from atopic and nonatopic (intrinsic) asthmatics. Journal of Immunology 1997;158:3539-44. Epub 1997/04/01.
- 34. Shahid SK, Kharitonov SA, Wilson NM, Bush A, Barnes PJ. Increased interleukin-4 and decreased interferon-gamma in exhaled breath condensate of children with asthma. American Journal of Respiratory and Critical Care Medicine 2002;165:1290-3. Epub 2002/05/07.
- 35. Humbert M, Durham SR, Kimmitt P, et al. Elevated expression of messenger ribonucleic acid encoding IL-13 in the bronchial mucosa of atopic and nonatopic subjects with asthma. The Journal of Allergy and Clinical Immunology 1997;99:657-65. Epub 1997/05/01.
- Kotsimbos TC, Ernst P, Hamid QA. Interleukin-13 and interleukin-4 are coexpressed in atopic asthma. Proc Assoc Am Physicians 1996;108:368-73. Epub 1996/09/01.
- Komai-Koma M, McKay A, Thomson L, et al. Immuno-regulatory cytokines in asthma: IL-15 and IL-13 in induced sputum. Clin Exp Allergy. 2001;31:1441-8. Epub 2001/10/10.
- Tang L, Lin HG, Chen BF. Association of IL-4 promoter polymorphisms with asthma: a meta-analysis. Genetics and Molecular Research: GMR 2014;13:1383-94.
- Heinzmann A, Mao XQ, Akaiwa M, et al. Genetic variants of IL-13 signalling and human asthma and atopy. Human Molecular Genetics 2000;9:549-59.
- 40. Nie W, Liu Y, Bian J, Li B, Xiu Q. Effects of polymorphisms -1112C/T and +2044A/G in interleukin-13 gene on asthma risk: a meta-analysis. PloS one 2013;8:e56065.
- Wang ZD, Lian D, Shen JL, et al. Association between the interleukin-4, interleukin-13 polymorphisms and asthma: a meta-analysis. Molecular Biology Reports 2013;40:1365-76.
- Corren J, Lemanske RF, Hanania NA, et al. Lebrikizumab treatment in adults with asthma. N Engl J Med 2011;365:1088-98. Epub 2011/08/05.
- 43. Bagnasco D, Ferrando M, Varricchi G, Passalacqua G, Canonica GW. A Critical Evaluation of Anti-IL-13 and Anti-IL-4 Strategies in Severe Asthma. International Archives of Allergy and Immunology 2016;170:122-31. Epub 2016/09/17.
- 44. Jiang H, Harris MB, Rothman P. IL-4/IL-13 signaling beyond JAK/STAT. The Journal of Allergy and Clinical Immunology 2000;105(6 Pt 1):1063-70.
- 45. Wenzel S, Castro M, Corren J, et al. Dupilumab efficacy and safety in adults with uncontrolled persistent asthma despite use of medium-to-high-dose inhaled corticosteroids plus

a long-acting beta2 agonist: a randomised double-blind placebo-controlled pivotal phase 2b dose-ranging trial. Lancet 2016;388(10039):31-44. Epub 2016/05/01.

- Rabe KF, Nair P, Brusselle G, et al. Efficacy and Safety of Dupilumab in Glucocorticoid-Dependent Severe Asthma. N Engl J Med 2018;378:2475-85. Epub 2018/05/22.
- Castro M, Corren J, Pavord ID, et al. Dupilumab Efficacy and Safety in Moderate-to-Severe Uncontrolled Asthma. N Engl J Med 2018;378:2486-96. Epub 2018/05/22.
- 48. Smith SG, Chen R, Kjarsgaard M, et al. Increased numbers of activated group 2 innate lymphoid cells in the airways of patients with severe asthma and persistent airway eosinophilia. The Journal of Allergy and Clinical Immunology 2016;137:75-86 e8.
- 49. Wood LJ, Sehmi R, Dorman S, et al. Allergen-induced increases in bone marrow T lymphocytes and interleukin-5 expression in subjects with asthma. American Journal of Respiratory and Critical Care Medicine 2002;166:883-9.
- Dorman SC, Efthimiadis A, Babirad I, et al. Sputum CD34+IL-5Ralpha+ cells increase after allergen: evidence for in situ eosinophilopoiesis. American Journal of Respiratory and Critical Care Medicine 2004;169:573-7.
- Park SW, Kim DJ, Chang HS, et al. Association of interleukin-5 and eotaxin with acute exacerbation of asthma. International Archives of Allergy and Immunology 2003;131:283-90.
- 52. Kabesch M, Depner M, Dahmen I, et al. Polymorphisms in eosinophil pathway genes, asthma and atopy. Allergy 2007;62:423-8.
- Molfino NA, Gossage D, Kolbeck R, Parker JM, Geba GP. Molecular and clinical rationale for therapeutic targeting of interleukin-5 and its receptor. Clin Exp Allergy 2012;42:712-37.
- 54. Varricchi G, Bagnasco D, Borriello F, Heffler E, Canonica GW. Interleukin-5 pathway inhibition in the treatment of eosinophilic respiratory disorders: evidence and unmet needs. Current Opinion in Allergy and Clinical Immunology 2016;16:186-200.
- 55. Pelaia C, Vatrella A, Bruni A, Terracciano R, Pelaia G. Benralizumab in the treatment of severe asthma: design, development and potential place in therapy. Drug Design, Development and Therapy 2018;12:619-28.
- Pelaia G, Vatrella A, Busceti MT, et al. Role of biologics in severe eosinophilic asthma - focus on reslizumab. Therapeutics and Clinical Risk Management 2016;12:1075-82.
- Matera MG, Calzetta L, Rinaldi B, Cazzola M. Pharmacokinetic/ pharmacodynamic drug evaluation of benralizumab for the treatment of asthma. Expert opinion on drug metabolism & toxicology. 2017;13:1007-13.
- Pelaia C, Calabrese C, Vatrella A, et al. Benralizumab: From the Basic Mechanism of Action to the Potential Use in the Biological Therapy of Severe Eosinophilic Asthma. BioMed Research International 2018;2018:4839230. Epub 2018/06/05.
- 59. Al-Ramli W, Prefontaine D, Chouiali F, et al. T(H)17-associated cytokines (IL-17A and IL-17F) in severe asthma. The Journal of Allergy and Clinical Immunology 2009;123:1185-7.
- Chesne J, Braza F, Mahay G, Brouard S, Aronica M, Magnan A. IL-17 in severe asthma. Where do we stand? American Journal of Respiratory and Critical Care Medicine 2014;190:1094-101.

Epub 2014/08/28.

- 61. Vittal R, Fan L, Greenspan DS, et al. IL-17 induces type V collagen overexpression and EMT via TGF-beta-dependent pathways in obliterative bronchiolitis. American journal of physiology Lung Cellular and Molecular Physiology 2013;304:L401-14. Epub 2012/12/25.
- 62. Ji X, Li J, Xu L, et al. IL4 and IL-17A provide a Th2/Th17-polarized inflammatory milieu in favor of TGF-beta1 to induce bronchial epithelial-mesenchymal transition (EMT). International Journal of Clinical and Experimental Pathology 2013;6:1481-92. Epub 2013/08/08.
- 63. Agache I, Ciobanu C, Agache C, Anghel M. Increased serum IL-17 is an independent risk factor for severe asthma. Respiratory Medicine 2010;104:1131-7.
- Linden A, Dahlen B. Interleukin-17 cytokine signalling in patients with asthma. The European Respiratory Journal 2014;44:1319-31.
- Ciprandi G, Cuppari C, Salpietro AM, et al. Serum IL-23 strongly and inversely correlates with FEV1 in asthmatic children. International Archives of Allergy and Immunology 2012;159:183-6.
- Fattahi F, Brandsma CA, Lodewijk M, et al. Atopy and Inhaled Corticosteroid Use Associate with Fewer IL-17+ Cells in Asthmatic Airways. PloS one 2016;11:e0161433.
- 67. Wilson RH, Whitehead GS, Nakano H, Free ME, Kolls JK, Cook DN. Allergic sensitization through the airway primes Th17dependent neutrophilia and airway hyperresponsiveness. American journal of Respiratory and Critical Care Medicine 2009;180:720-30.
- Wakashin H, Hirose K, Maezawa Y, et al. IL-23 and Th17 cells enhance Th2-cell-mediated eosinophilic airway inflammation in mice. American journal of Respiratory and Critical Care Medicine 2008;178:1023-32.
- 69. Bajoriuniene I, Malakauskas K, Lavinskiene S, et al. Response of peripheral blood Th17 cells to inhaled Dermatophagoides pteronyssinus in patients with allergic rhinitis and asthma. Lung 2012;190:487-95.
- 70. Camargo LDN, Righetti RF, Aristoteles L, et al. Effects of Anti-IL-17 on Inflammation, Remodeling, and Oxidative Stress in an Experimental Model of Asthma Exacerbated by LPS. Frontiers in Immunology 2017;8:1835. Epub 2018/01/31.
- Zhu M, Wang T, Chen R, Wang C, Liu S, Ji Y. Association between interleukin-17a gene polymorphisms and asthma risk: a metaanalysis. Asian Pacific Journal of Allergy and Immunology 2016;34:115-23.
- 72. Busse WW, Holgate S, Kerwin E, et al. Randomized, double-blind, placebo-controlled study of brodalumab, a human anti-IL-17 receptor monoclonal antibody, in moderate to severe asthma. American journal of Respiratory and Critical Care Medicine 2013;188:1294-302. Epub 2013/11/10.
- Abbas AK, Murphy KM, Sher A. Functional diversity of helper T lymphocytes. Nature 1996;383(6603):787-93.
- 74. Cosmi L, Maggi L, Santarlasci V, et al. Identification of a novel subset of human circulating memory CD4(+) T cells that produce both IL-17A and IL-4. The Journal of Allergy and Clinical Immunology 2010;125:222-30 e1-4.

- 75. Zhou L, Chong MM, Littman DR. Plasticity of CD4+ T cell lineage differentiation. Immunity 2009;30:646-55.
- 76. O'Shea JJ, Paul WE. Mechanisms underlying lineage commitment and plasticity of helper CD4+ T cells. Science 2010;327(5969):1098-102.
- 77. Murphy KM, Stockinger B. Effector T cell plasticity: flexibility in the face of changing circumstances. Nature immunology 2010;11:674-80.
- 78. Jameson SC, Masopust D. Diversity in T cell memory: an em-

barrassment of riches. Immunity 2009;31:859-71.

- 79. Locksley RM. Nine lives: plasticity among T helper cell subsets. The Journal of Experimental Medicine 2009;206:1643-6.
- 80. Fahy JV. Eosinophilic and neutrophilic inflammation in asthma: insights from clinical studies. Proceedings of the American Thoracic Society 2009;6:256-9.
- Choy DF, Hart KM, Borthwick LA, et al. TH2 and TH17 inflammatory pathways are reciprocally regulated in asthma. Science Translational Medicine 2015;7(301):301ra129. Epub 2015/08/21.